

## Center for Musculoskeletal Research

Massachusetts General Hospital  
50 Blossom Street, Thier 10 & 11  
Boston, MA 02114-2696

Email: CSRMAIL@PARTNERS.ORG

Website: www.csr-mgh.org

## Histology and Histomorphometry Core

Email: endocrinehistocore@partners.org

### **Protocol for IHC of phosphoERK in growth plate (with tyramide amplification, Perkin Elmer, Cat. No. NEL700)**

- Controls:**
1. no 1<sup>st</sup> Ab, or 1:100 nonspecific IgG
  2. no amplification

**Preparation of reagents:**

1. 90% methanol, 10% DMSO (for 200ml need 180ml methanol, 20ml DMSO)
2. 11 Ix PBS
3. 0.3% H<sub>2</sub>O<sub>2</sub> (for 200ml need 2ml 30% H<sub>2</sub>O<sub>2</sub> in 200ml methanol)
4. Biotinyl Tyramide: add DMSO (0.3 ml [NEL700A] or 1.2 ml [NEL700]), store at 4C (stable 6 months)
5. Biotinyl Tyramide Working Solution: dilute stock 1:50 with Ix Amplification Diluent, 100ul/slide
6. Wash Buffer (TNT; 0.1M Tris, 0.15M NaCl, 0.05% Tween, pH 7.5): for 31 need 300ml 1M Tris, pH 7.5, 90ml 5M NaCl, 1.5ml Tween, 2610ml water
7. Blocking Buffer (TNB; 0.1M Tris, 0.15M NaCl, 0.5% Blocking Reagent, pH 7.5): for 10ml need 1ml 1M Tris, 0.3ml 5M NaCl, 0.05g Blocking Reagent, heat to 60C to completely dissolve Blocking Reagent, can be stored at -20C in aliquots, at RT no longer than 24h
8. SA-HRP (1:100 in TNB buffer)
9. DAB kit for visualization

**Procedural notes:**

1. need humidified chamber (slide box with damp paper towels) for ALL incubation steps
2. staining tubs for incubations need volumes of 200ml

**Protocol:**

1. heat paraffin slides for 30min at 55C to melt wax
2. deparaffinize slides in 2 changes of xylene x 5min
3. rehydrate slides in series of graded alcohol 100%, 80%, 70% x 5 min
4. incubate slides in 90% methanol, 10% DMSO for 20min at RT
5. wash with PBS
6. incubate with 0.3% H<sub>2</sub>O<sub>2</sub> in methanol for 20min
7. wash with PBS
8. block slides with 100ul TNB for 30min in humidified chamber at RT, drain off
9. apply 100 ul of 1<sup>st</sup> Ab anti-phosphoERK in TNB buffer, Cell Signaling, Cat.No. 4377S) for overnight at 4 °C..

10. wash 3x5min with TNT buffer at RT with agitation
11. apply 100ul 2nd biotinylated Ab (1:300 anti-rabbit biotinylated IgG in TNB, Vector, Cat.No. BA1000) for 1h at RT
12. wash 3x5min with TNT buffer
13. apply 100ul SA-HRP (diluted 1:100 in TNB) for 30min at RT
14. wash 3x5min with TNT buffer
15. apply 100ul Biotinyl Tyramide Working Solution for 5 min at RT

16. wash 3x5min with TNT buffer
17. add 100 ul of SA-HRP and incubate 30min at RT
18. wash 3x5min with TNT buffer
19. visualize with chromogenic substrate 5min (time varies) in
  - 5mlH<sub>2</sub>O
  - 4 drops DAB
  - 2 drops buffer
  - 2 drops H<sub>2</sub>O<sub>2</sub>
  - 2 drops NiCl (optional: will give grey/black stain as compared to brown with DAB only)
20. wash with distilled water until desired signal intensity achieved (1-3min)
21. counterstain with hematoxylin (5sec; Histo-core sequence) or toluidine blue (see separate protocol)